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Silter cheese, a traditional Italian dairy product: a source of feasible probiotic strains

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9 **KEY WORDS:** Silter, cheese, probiotics, food

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12 **ABSTRACT**

13 Silter cheese is a traditional hard cheese, produced in Valcamonica, Brescia, Italy. A total of 426
14 lactic strains isolated from Silter were analysed to determine their probiotic characteristics. 274
15 strains out of 426 were found to produce bacteriocins against at least one of eight different
16 pathogens (*Salmonella enterica*, *Listeria monocytogenes*, *Salmonella derby*, *Salmonella*
17 *thyphimurium*, *Salmonella napoli*, *Staphylococcus aureus*, *E. coli* O157:H7, *Salmonella*
18 *enteritidis*). In addition, 211 of 274 bacteriocin-producer strains adhered to Caco-2 cells and were
19 characterized by RiboPrinter, revealing predominance of *Enterococcus faecalis* (26%) and
20 *Enterococcus durans-faecium* (22%). These findings suggest that Silter may qualify as an
21 important source of feasible probiotic strains.

22

23 INTRODUCTION

24 Functional foods are foods that provide health benefits beyond basic nutrition due to certain
25 physiologically active components. A definition of functional foods was first depicted in Japan in
26 the 1980s, referring to processed foods containing ingredients that aid specific bodily functions.
27 Health benefits related to consumption of functional food are mostly triggered by probiotic
28 microorganisms, such as live lactobacilli and bifidobacteria [1].

29 The interest in functional foods is growing rapidly, driven by several factors including the rapid
30 advances in scientific knowledge, consumer demand for healthier foods, the aging populations,
31 the ever-increasing health-care costs, and the food industry desire to fulfil the consumer appetite
32 for products derived from food that can promote good health condition [2]. In the category of
33 nutraceuticals, comprised of functional foods and supplement components, the role of probiotics
34 appears to be even more significant. The term “probiotics” was first used by Parker in 1974 [3]
35 to define organisms or substances able to contribute to intestinal microbial balance [3].
36 Schrezenmeir and de Vrese [4] defined probiotics as “a preparation of or a product containing
37 viable, microorganisms in sufficient numbers which, by implantation or colonization, alter the
38 microflora in a compartment of the host and by that exert beneficial health effects in the host”.
39 Probiotics are largely used in fermented dairy products and cheeses [5]. The Japanese
40 Association of Fermented Milks and Fermented Milks Drinks has established a minimum level
41 of 1×10^7 viable lactic-acid-producing-bacteria (LAB) per ml for probiotic milk product [6].
42 Finally, the FAO-WHO guidelines define probiotics as “live microorganisms which, when
43 administered in adequate amount, confer health benefits to the host” [7].

44 The beneficial effects of probiotics are not limited to the large intestine, since they also extend to
45 the nasal cavities and to the urogenital system [8]. Probiotics related health benefits include
46 improvement of intestinal peristalsis, decrease in cholesterol level and enhancement of immune
47 system activities. Adhesion to intestinal cells and competitive activities against potential
48 pathogens or dismetabolic microbial populations are significant/beneficial features of lactic
49 bacteria [9, 10]. Functional foods enriched with probiotics are highly popular among different
50 European consumers' categories, and they are largely available on the market. In spite of their
51 increasing economic relevance, functional foods are not specifically regulated by European
52 legislation [11]. The evolution of legislation on the matter is very slow; currently only Japan, the
53 U.K., the U.S.A., and Scandinavian countries have achieved some considerable progress.
54 Moreover, the labelling of functional foods is far from being informative, and only provides
55 scanty information about nutritional value, storage, and cooking recipes [12].

56 Silter cheese is a PDO (Protected Designation of Origin) hard or semi-hard cheese, produced
57 using milk from Italian Brown and Italian Red Pied cows in the province of Brescia, northern
58 Italy, since the end of the 16th century as found in a document written by a local chancellor
59 [13]. Its name derives from the local dialect and refers to the shelter and dairy used by humans
60 and animals in high Alps mountains pastures. Silter cheese is made in the shape of a cylinder of
61 30 to 40 cm in diameter and 8 to 12 cm in height, with a weight ranges from 6 to 15 kg. The
62 crust is yellow, hard and smooth. The yellow straw inner part presents many little holes and the
63 flavour is slightly sweet and aromatic (Figure 1). This characteristic derives from typical
64 aromatic grass growing at high altitudes which is used to feed the cows. One year's seasoning

65 makes the holes grow in number, the flavour and aroma more intense, and the cheese ready for
66 grating.

67 We studied the probiotic properties of the micro flora found in Silter cheese. LAB isolated from
68 Silter cheese were characterised both genetically and phenotypically. The data gathered from this
69 preliminary probiotic screening was used to define a baseline to be used for the adoption of
70 microbiological standards in the qualitative assessment of cheese production.

71 **MATERIALS AND METHODS**

72 *Samples*

73 Silter is a hard/semi-hard traditional cheese produced in northern Italy since the end of the 16th
74 century. Silter cheese production process: (i) Raw, partially skimmed milk from the two daily
75 milkings is warmed to 35-36 °C; after (ii) adding the rennet, the natural whey from previous
76 work, and mesophilic and termophilic starters, it coagulates in around 40 – 45 minutes; (iii) the
77 curd which has formed is then broken using a traditional tool called *spada* or *spannarola*; it rests
78 into a vat for 10 – 40 minutes; and then, using a tool called *spino*, it is broken down again into a
79 rice sized pieces and cooked at 46 – 48 °C for 10 – 15 minutes; (iv) curd rests for nearly 1 hour
80 in vat at 45-46 °C; after that it is transferred into a marked mould and dry salted for 2 days. The
81 cheese is seasoned for at least 200 days. Following the guidelines of the European Commission
82 Regulation (EC) No. 2081/92 [14], in 2005-2007 a project aimed at the study of the
83 microbiological features of Silter cheese was carried out, helping this cheese to be awarded with
84 PDO designation. A total of 50 samples of Silter cheese, seasoned for 200 days, having a mean

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85 weight of 500 g, were obtained from 50 different cheese manufactured in two of the around forty
86 different cheese industries members of the consortium for protection of Silter cheese in
87 Lombardy, Italy. Samples were transported to the laboratory at ≤ 4 °C and on arrival their
88 interior portion was analyzed after being cut by means of a sterile knife.

89 *Strains isolation.*

90 Lactic acid bacteria strains were isolated onto MRS agar (Man Rogosa Sharpe) (Oxoid) [15, 16].
91 The isolates obtained from MRS agar were cultured in MRS broth (Oxoid) at 37 °C [17] and
92 titrating $\geq 1 \times 10^7$ colonies/ml were stored at -80 °C with 20% glycerol.

93 *Bacteriocins production.*

94 Production of bacteriocins was assessed against 8 different pathogens: *Listeria monocytogenes*
95 (ATCC 19115); *Salmonella enterica* (IZSLER 230747); *Salmonella derby* (IZSLER 259473);
96 *Salmonella thyphimurium* (ATCC 6994); *Salmonella napoli* (IZSLER 337144); *Salmonella*
97 *enteritidis* (IZSLER 240752); *Escherichia coli* O157:H7 (IZSLER 675); *Staphylococcus aureus*
98 (IZSLER 542). The productions of bacteriocins were assessed by a well diffusion assay as
99 described previously [16, 18, 19]. Briefly, the plate wells were soaked with a suspension of lactic
100 strain cultured in MRS broth at 37 °C for 24 hours, while the control plate was loaded with 10
101 $\mu\text{g/ml}$ of Nisin (Sigma-Aldrich) (pH 8) with addition of EDTA for analysis of Gram positive
102 pathogens. The plates were incubated at 30 °C for 5 hours, covered with a layer of PCA (Plate
103 Count Agar) (Oxoid) and of PCA mixed with a broth suspension of the pathogen (grown at 37
104 °C in Brain Heart Infusion broth) , and then incubated overnight at 37 °C.

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105 *In vitro adhesion assay.*

106 The lactic acid bacteria strains able to produce bacteriocins were tested by an *in vitro* adhesion
107 assay on Caco-2 cells line, originally isolated from a human colon adeno-carcinoma, as
108 described previously [20]. For the adhesion assay, monolayers of cells, cultured in Dulbecco
109 modified Eagle's minimal essential medium, were prepared in 6 well tissue culture plates. Forty-
110 eight-hours cultures (10 ml) in MRS broth of each lactic bacterium isolate were harvested and
111 washed in physiological saline. Tenfold serial dilution in physiological saline were inoculated on
112 differentiated Caco-2 cells monolayers, after removing the growth medium. After a contact of
113 1.5 hours at 37 °C in 5% CO₂ atmosphere, the cells were washed twice using physiological
114 saline, trypsinised and re-suspended in 4 ml of physiological saline with addition of 500 µl of
115 foetal bovine serum. One ml of each cell suspension was mixed with 13 ml of MRS agar and the
116 plates were incubated at 37 °C in 5% CO₂ for 3 days. The colonies grown in the plates were
117 counted to verify bacterial adhesivity [16]. A two logarithms difference in bacterial count before
118 and after contact with Caco-2 culture cells was used to evaluate bacterial adhesivity.

119 *Strains characterisation.*

120 All the lactic acid bacteria strains able to produce bacteriocins and to adhere to Caco-2 cells were
121 further characterised by automated ribotyping using the RiboPrinter system (DuPont-
122 QualiconLtd, Wilmington, Del.) as described previously [21, 22].

123

124 **RESULTS AND DISCUSSION**

125 A total of 426 strains of lactic-acid-producing bacteria at concentration $\geq 1 * 10^7$ colonies/ml
126 were isolated from the Silter cheese samples. This value has been proposed as a distinctive
127 criterion for probiotic products [6]. Noteworthy, in Silter cheese, fulfilment of this criterion is
128 not derived from production artefacts, as the presence of lactic acid bacteria is not due to
129 addition during production, but it is rather an intrinsic property of the traditional techniques
130 inherited over more than four centuries of history.

131 Adhesion to intestinal cells and production of factors inhibiting potential pathogens or
132 dismetabolic microbial populations are significant mechanisms by which probiotics exert
133 beneficial effects on health. These properties can be investigated *in vitro* in order to assess the
134 probiotic qualities of lactic acid producing bacteria. In this study, out of 426 bacterial isolates, a
135 total of 242 strains (56,8%) produced bacteriocins against *Listeria monocytogenes*, 251 (58,9%)
136 against *Salmonella enterica*, 251 (58,9%) against *Salmonella derby*, 244 (57,2%) against
137 *Salmonella thyphimurium*, 253 (59,3%) against *Salmonella napolis*, 241 (56,5%) against
138 *Salmonella enteritidis*, 249 (58,4%) against *Escherichia coli* O157:H7 and 247 (57,9%) against
139 *Staphylococcus aureus* (Figure 2). Production of bacteriocins against, at least, one of the 8
140 bacterial pathogens was demonstrated by the agar well diffusion assay in 274 out of 426 (64%)
141 isolates obtained from Silter cheese. An interesting result is that most of the 274 bacteriocins-
142 positive isolates exhibited a broad spectrum of inhibition activity against both Gram-negative
143 and Gram-positive bacterial pathogens. The mechanisms by which bacteriocins inhibit growth of
144 bacterial pathogens are unclear. Abee et al. [23] demonstrated that the inhibitory activity of these

145 substances is limited to Gram-positive bacteria. On the other hand, Stevens *et al.* [24] and Ray
146 [25] demonstrated that bacteriocins are also active against *Salmonella* spp. and other Gram-
147 negative pathogens.

148 Two hundred and eleven out of 274 bacteriocins-producing isolates (77%) were also found to be
149 able to adhere to Caco-2 cells by an *in vitro* adhesion assay. The ability to adhere to Caco-2 cells
150 is a good proxy to assess the ability of the bacteria to adhere to human epithelial cells in the
151 intestinal tract [9, 10]. Adhesion to intestinal cells and competition with potential pathogens or
152 dismetabolic microbial populations are believed to be an important mechanism by which lactic
153 bacteria exert their beneficial activity.

154 Accordingly, a total of 211 lactic acid bacteria strains isolated from Silter cheese displayed
155 probiotic characteristics (production of bacteriocins and ability to adhere to Caco-2 cells). These
156 strains were further characterized by ribotyping, revealing an highly heterogeneous microflora
157 population. The predominant strains were *Enterococcus faecalis* (26%) and *Enterococcus durans*
158 *faecium* (22%), followed by *Lactobacillus curvatus* (8%), *Lactobacillus pantheris* (8%),
159 *Lactobacillus lactis* (7%), *Lactobacillus salivarius* (7%), *Lactobacillus paraplantarum* (5%),
160 *Streptococcus thermophilus* (5%), *Enterococcus* spp. (4%), *Lactococcus garviae/Enterococcus*
161 (4%), *Lactobacillus delbrueckii bulgaricus* (2%) and *Lactobacillus parabuchneri* (2%).
162 *Enterococci* (48% of isolated LAB in this work) are natural members of the intestinal flora in the
163 large bowel of humans and in the intestinal tract of mammals and birds [26]. They are also found
164 in soil, plants, and water. *Enterococci* have emerged as a major cause of nosocomial infections,
165 and, within this group, *Enterococcus faecalis* causes the majority of human enterococcal

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166 infections. These bacteria may be associated with bacteremia, systemic or local infections in the
167 urinary tract, abdomen, wounds and endocardium [27]. However, *Enterococci* have also been
168 studied for their possible use as probiotics. Administration of an *E. faecalis* strain has been
169 shown to decrease diarrhea [28].

170 In this study, *Lactobacillus* spp. accounted for nearly 40% of the isolates made from Silter
171 cheese. Lactobacilli are an important part of the human flora, but they can be pathogenic under
172 certain conditions. *Lactobacillus* species are used for production of fermented food, such as
173 yogurt, cheese, beer, wine, cider, chocolate, and animal feeds, such as silage. In recent years,
174 interest has increased interest in their use as probiotics and many probiotic formulations contain
175 *Lactobacillus* species. Various species of lactobacilli may be beneficial in certain infectious
176 diarrheas or other diseases [29].

177 Silter cheese has peculiar characteristics deriving from the use of local ingredients and
178 production techniques developed over centuries of traditions and linked to the Valcamonica
179 territories. This cheese is considered a territorial heritage and the techniques/modalities of
180 production have recently been codified and disciplined according to PDO guidelines [14] in
181 order to guarantee food safety and preserve the identity of this cheese. The findings of this study
182 provide evidence that this traditional food possesses a lactic flora with functional activity. The
183 potential probiotic activity, along with the nutritional properties and the territorial specificity,
184 surely adds intrinsic value to Silter cheese and this could be a claim in advertising and marketing
185 promotions, respecting the requirements of the European community legislation [29].

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186 In conclusion, in this study many information were gathered on the microflora of a traditional
187 Italian processed food. Based on these findings, Silter cheese represents an important source of
188 probiotic strains, but it is not still possible to define it as a functional food, because this should be
189 further evaluated, by repeating and extending the investigations. It should also be assessed
190 whether, and, in case, to which extent, virulence and drug resistance factors are harbored in the
191 lactic microflora of Silter cheese, especially in *Enterococcus* genus. *Enterococci* need to be
192 considered safe to confer probiotic activity to the matrix to which they belong. These aspects are
193 not commonly considered when assessing the microbiological features of food products. The
194 above mentioned information could be used to support and promote this category of products on
195 both domestic and international markets.

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